

S. E. Luria and M. Delbruck (1943). Mutations of bacteria from virus sensitivity to virus resistance. *Genetics* 28:491

[Presented by: Steve Carr (scarr@mun.ca), 14 January 2016]

Background

[Max Delbruck](#) (1906 -1981) & [Salvador Luria](#) (1912 - 1991)
[1969 Nobel Prize](#) in Physiology or Medicine

Bacteriology <1940 not influenced by genetic thinking

No nuclei: do they have "genes"?

No individual "phenotypes": colonies of 10^6 s of bacteria simultaneously

No sex: genetic crosses not possible

[Discovery of bacterial sex led to [1958 Nobel Prize](#)]

bacteriophages ("**phages**") - "*Subcellular parasites that infect, multiply within, & kill bacteria.*"

T1 phages are active on [E. coli](#)

[phage] >> [bacteria] \Rightarrow no bacterial colonies grow: bacteria are **Ton^S** ("T-one sensitive")

[phage] ~ [bacteria] \Rightarrow some bacterial colonies grow: bacteria are **Ton^R** ("T-one resistant")

Ton^R phenotype is **stable, heritable**

all descendant bacteria are **Ton^R**

phenotype *persists* in the *absence* of T1

Two Hypotheses (d'Herelle 1926 vs Brunet 1929)

1. **Ton^R phenotype** **induced** by exposure of bacteria to phage

Each bacterium has small, finite prob. of survival ($\sim 1 / 10^7$);

Survivors have altered *metabolic phenotype*, transmitted to offspring:

changed *phenotype* persists in *genotype*

Bacteria *adapt* to their environment :

a **Lamarckian** hypothesis: **inheritance of acquired characteristic**

2. **Ton^R phenotype** occurs **spontaneously**, prior to exposure of bacteria to phage

Some rare bacteria ($\sim 1 / 10^7$) are already **Ton^R**

have undergone **genetic mutation** to a stable *genotype*

changed *genotype* regenerates *phenotype*

a **Darwinian** hypothesis: **Ton^R bacteria** are **selected**

Materials & Methods

Hypotheses predict *different* distributions of **Ton^R** phenotypes among cultures

Induction Hypothesis predicts: **$n / N = a$**

where **n** = number of **Ton^R** bacteria observed out of

N = number of **Ton^S** bacteria plated, where

a = **probability of conversion** from **Ton^S** to **Ton^R**

Then, **n** should be *constant* wrt **N**

Mutation Hypothesis predicts: **$n / N = ga^{2^g} / 2^g = ga$**

where **a** = **mutation rate** (# mutations / cell / generation)

g = # **generations** to go from **1** \rightarrow **N** bacteria, so that

N = **2^g** doublings occur, of which

n = **ga^{2^g}** produce mutant **Ton^R** bacteria

because a mutation in the **i** th generation contributes **a^{2ⁱ}2^{g-i}** = **a^{2^g}** mutants

Then, **n** should *increase* wrt **N** as **g** increases

How can differences in **n** be evaluated?

Thought experiment:

Consider four cultures each started from a single bacterium

after **g = 4** generations, expect **64 cells** from **60 divisions**

plate each culture separately w/ **T1**, count total # **Ton^R**

Suppose **10 Ton^R** colonies observed: what distribution (**variance**, "*fluctuation*") expected?

Induction Hypothesis:

Ton^r induction occurs in *last* generation upon exposure to T1

probability of induction (a) is *uniform* / bacterium

a = **10 inductions** / **64 cells** = 15%

observe = **3, 1, 5, & 1** Ton^r colonies

mean = $10 / 4 = 2.5 \text{Ton}^r$ per culture

variance = **2.75**

Expect **Poisson Distribution** for rare, random events: **variance = mean**

Homework: Evaluate **16 plants / 64 quadrats** distribution by **Chi-Square**

Mutation Hypothesis

Ton^r mutations occur spontaneously, *prior* to exposure to T1

mutation rate (a) = **2 events** / **60 cell divisions** = **0.033** mutations / cell / generation

mean = $(2 + 0 + 8 + 0) / 4 = 2.5 \text{Ton}^r$ as before

After 4 generations, early **mutations** leave more offspring

variance = **10.75**

after 5 generations, # of Ton^r cells *doubles* in each culture:

variance = **48.00**

Mutation Hypothesis predicts variance >> mean, as g increases

Experimental procedure:

"The first experiment was done on the following Sunday morning.

(In a letter dated January 21 [1943], Delbruck exhorted me to go to church"

Twenty x **200 ul "individual cultures"**

One x **10 ml "bulk culture"**

Inoculate with ~ **10^3 bacteria** @

Grow for **g = 17** generations

⇒ ~ 10^8 bacteria / ml

Plate entire "individual cultures"

& 200 ul aliquots of "bulk culture" on petri dish w/ T1

Results

	<u>Bulk Cultures</u>	<u>Individual Cultures</u>
Experiment ##	10a	16
Mean	16.7	11.3
Variance	15	694

Single bulk culture (e.g., **Experiment 10a**):

a = $n / N = (16.7 / (0.2 \text{ ml} \times 10^8 \text{ bacteria / ml})) = 8 \times 10^{-7}$ variants / cell

variance ~ mean ⇒ random distribution

Expected result for either induced or spontaneous hypotheses: a **control**

Multiple individual cultures (e.g., **Experiment 16**):

mean ~ mean in bulk

variance >> variance in bulk:

Experiment supports Mutation Hypothesis !

Calculation of **Mutation rate** (a)

mean # mutations / culture = **aN**

Poisson predicts **null class** = $p_0 = e^{(-a/N)}$

where p_0 = fraction of cultures with **no** Ton^r mutants

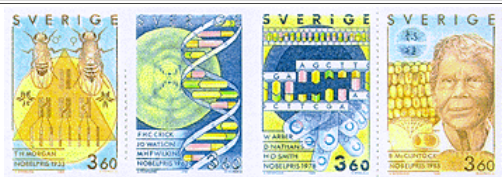
Rewrite as **a = -ln(p_0 / N)**

$p_0 = 11 / 20 = 0.55$ from **Experiment 16**

$N = 0.2 \text{ ml} \times 10^8 \text{ bacteria / ml}$

Then **a = -ln 0.55 / (0.2 x 10^8) = 3×10^{-8}** mutations / cell / generation

Conclusion



January 24, 1943

Salvador

"You are right about the difference in fluctuations of resistants, when plating samples from one or from several cultures. In the latter case, the number of clones has a Poisson distribution. I think what this problem needs is a worked out and written down theory, and I have begun doing so."

Max

The MS of the theory arrived on February 3rd"

Luria on the significance of these experiments:

- (1) **"Adequate evidence" of spontaneous mutation** as source of genetic variation
- (2) Provided **method** for measuring **mutation rates**, and therefore is
- (3) **"The Birth of Molecular Genetics"**
bacteria can be used to measure **extremely low gene mutation rates**

Homework: repeat all statistical calculations for **Experiments 3 & 21a**

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